

RiverWatch Water Quality Volunteer Monitoring Program

The Ipswich River Watershed Association (IRWA) is the voice of the Ipswich River. IRWA works to protect nature and make sure that there is enough clean water for people, fish and wildlife, today and for our children and theirs.

2014 Annual Results Report

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Executive Summary

In 1997 the Ipswich River was listed as one of the 20 most threatened rivers in America. The level of threat to the Ipswich River was heightened in 2003 when it was ranked the third most endangered river in America by American Rivers, a national nonprofit, primarily due to low flow problems (American Rivers 1997 and 2003, IRWA 2003).

Much of the upper half of the River dried up or was reduced to isolated stagnant pools in the summers of 1995, 1997, 1999, 2001, 2002, 2003, and 2005. In 1999, the River experienced record low-flows in May, June, July and August. In 2000, the United States Geological Survey (USGS) completed a model of river flow that linked withdrawals for regional water supply with low flows in the Ipswich River. Major fish kills were also documented in 1995, 1997, 1999, 2002, and 2005.

Low flows continue to be a threat to the Ipswich River. In order to assess the health of the Ipswich River, the Ipswich River Watershed Association has maintained the RiverWatch Volunteer Water Quality Monitoring Program since 1997. Volunteers collect data monthly from March-December on weather conditions, rain in the last 48 hours, water color, water odor, water temperature, dissolved oxygen, velocity, depth, cross-sections and conductivity. Streamflow is also monitored at two sites in the upper watershed, where consistent flow data has been lacking. In 2014, volunteers monitored a total of 32 sites monthly from March to December. Three sites were monitored for streamflow with dedicated staff gages and nine sites were sampled for benthic macroinvertebrates. Benthic Macroinvertebrate monitoring is covered in a separate report.

Results

The Ipswich River and many of its tributaries continue to show impairment for dissolved oxygen and flow and relative abundance of organisms. Dissolved oxygen (DO) is necessary for all forms of life that depend on the river. DO is influenced by many factors including flow and temperature. Dissolved oxygen levels below 5 mg/L create a stressful environment for fish and other aquatic organisms. Levels below 3 mg/L can be fatal to organisms that cannot move to areas of higher concentration. Large fish kills can result from DO levels that fall below 1-2 mg/L, even if those levels are present for only a few hours. Certain fish species, like brook trout, are especially sensitive to low DO.

Low DO conditions have been widespread and frequent since monitoring began in 1997. In 2014, 34% of the collected samples did not meet the state standard for dissolved oxygen concentration of 5 mg/L for class B waters. Figure 1 illustrates average summer dissolved oxygen concentration values at all sites. Sites located in the upper section of watershed continue to show a higher degree of impairment for dissolved oxygen than sites elsewhere. The upper watershed includes the towns of Wilmington and North Reading in the southwestern area of the outlined watershed in figure 1.

All temperature samples met Massachusetts State Water Quality Standards. This indicates that temperatures are in an acceptable range along the Ipswich River. This may be an indicator of the importance that cool groundwater plays in providing the river's baseflow in summer. Shading from trees along the river also benefits water temperature. It is important to note that this measure does not consider the most extreme conditions as temperatures cannot be recorded when there is little (or no) water present in the river during extreme low flows. Also, monitoring is conducted in the morning, and may

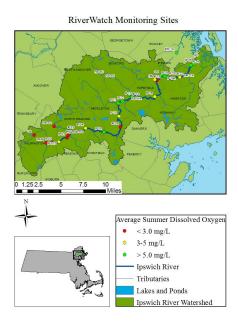


Figure 1. Average summer dissolved oxygen levels for 2013. Sites in red (< 3mg/L) represent a highly stressed environment for fish and other aquatic organisms.

not represent the highest temperatures that occur in the course of that day or month.

There must be water in the river for most aquatic organisms to survive. The Ipswich River experiences significant periods of extreme low flow during many years. Withdrawals for drinking and irrigation water are the primary cause of unnaturally low flows in the Ipswich River (Armstrong 2001, Zarrielo and Ries 2000). Low flows can be expected to occur seasonally, but the low flows observed in the Ipswich River are about 1/10th of what is considered "natural" (Zariello and Ries, 2000). Due to low flow, the Ipswich River is classified as highly stressed by the MA Water Resources Commission (2001) and impaired under section 303(d) of the Clean Water Act.

Streamflow gauges maintained by the United States Geological Survey (USGS) have recorded regular episodes of extended extreme low flow events since monitoring began in 1997. "Extreme low flow" is defined based on the USGS summer "ecological protection flow" (Horsley and Witten 2002), that "provides adequate habitat for the protection of fisheries" (Ibid). Extreme low flows were observed for 93 days in 2014, primarily during the summer and early fall.

Conductivity measures the ability of water to pass an electrical current resulting from the presence of dissolved solids (or salts) such as chloride, sulfate, sodium and calcium. Significant changes in conductivity can be an indicator that a discharge or some other source of pollution has entered the water. Rivers that can support healthy fisheries should be in the range of 150 to 500

 μ S/cm. In 2014, conductivity levels greater than 500 μ S/cm were recorded at 22% of sites, with many of these sites concentrated in the upper watershed. Elevated readings recorded in March and April may be the result of road-salt applications from the winter entering the river through stormwater runoff events. Elevated readings during the summer may be an indicator of other sources such as septic systems.

Conclusion

The upper watershed continues to experience low dissolved oxygen levels, especially during the summer months, despite low flow conditions not being as severe as in years prior to 2006 when the town of Reading discontinued using wells adjacent to the Ipswich River. Martins Brook continues to experience severe low flows near active groundwater wells. Low flows impact the biological health of the watershed.

The primary cause of impairments in the Ipswich River watershed are low flow alterations due to water withdrawals and impervious surfaces contributing to stormwater runoff. Under these conditions, dissolved oxygen levels decrease below what is suitable to aquatic life such as fish and macroinvertebrates that are an important part of the aquatic food web.

Water has remained in the river year-round since Reading discontinued well use, showing that reductions in water withdrawals and water restrictions by towns can have a beneficial effect on the Ipswich River.

Our deepest thanks to our volunteers that have monitored on sunny and rainy days, in cold and heat and high and low river flows. Thank you for your considerable efforts and dedication to the Ipswich River!

Section 1: Overview of the RiverWatch Monitoring Program

1.1 Description

The Ipswich River Watershed Association has conducted the RiverWatch water quality monitoring program since 1997. The program enlists a group of volunteers to collect water quality data on the Ipswich River and its tributaries. The purpose of the program is to establish baseline data in order to identify and address impairments to water quality and quantity, as well as to promote awareness and stewardship of the river. The RiverWatch program expanded upon an earlier, informal water quality monitoring program that ran from 1988 – 1996. An EPA-approved Quality Assurance Project Plan (QUAPP) was finalized in 1999 and most recently updated and approved by MassDEP in 2013. The goal of the RiverWatch program is to provide high quality data regarding the health of the Ipswich River. This monitoring program has established a crucial baseline of water quality and biological data, which continues to enable IRWA to work with researchers and government officials to better manage the watershed and improve the condition of the Ipswich River.

The specific goals of regularly monitoring the Ipswich River and its tributaries include:

- Defining the baseline water quality conditions of the Ipswich River and key tributaries.
- Defining the range of dissolved oxygen concentrations, temperature and conductivity over the range of annual conditions in both mainstem and tributary locations.
- Determining the relative water level and flow at a variety of ungauged locations around the basin.
- To observe the River, habitat and wildlife, and report on observations.
- To identify pollution hotspots.
- To educate watershed residents about the river.
- To promote stewardship of the river.
- Inform ongoing restoration efforts

Monitors collect data monthly on weather conditions, rain in the last 48 hours, water color, water odor, water temperature, dissolved oxygen, conductivity, velocity and depth. Streamflow data is recorded at two official gaging stations maintained by the USGS. Streamflow is also monitored at three additional sites established in 2012 in cooperation with the Massachusetts Division of Ecological Restoration.

The purpose of this report is to summarize data collected in 2014 by volunteers for the RiverWatch program. Specific site data are available in the appendix.

Data collected by IRWA will be reported to IRWA members, state agencies, interested organizations, and conservation commissions through reports and presentations on the collected data. Atypical data will be reported to the appropriate agencies. Atypical data include dissolved oxygen data that vary significantly from adjacent sites over one or more months. Extended periods of no flow or extremely low dissolved oxygen (less than 2 mg/L) are also considered

extremely important and will be presented to state agencies. When dissolved oxygen levels fall below 2 mg/L the health of fish and other aquatic organisms can be severely impacted.

Section 2: An Introduction to the Ipswich River

The Ipswich River watershed is 155 square miles and includes all or part of 21 communities in northeastern Massachusetts. The topography of this Atlantic coastal plain basin is characterized by low relief, with an average grade of 3.1 feet per mile. The length of the river is a meandering 40 miles. The surficial geology of the region consists primarily of glacial till with stratified sand and gravel deposits covering about 43 percent of the basin and alluvial deposits covering about 3 percent of the basin. Extensive wetlands are present along the River and streams within the Ipswich River basin. These wetlands protect surrounding areas during flooding as well as positively affect the water quality of the River and streams in the basin.

This river system supplies water to more than 330,000 people and thousands of businesses, providing all or part of the water supply for 14 communities: Beverly, Danvers, Hamilton, Ipswich, Lynn, Lynnfield, Middleton, North Reading, Peabody, Salem, Topsfield, Wenham, and Wilmington.

In 1997 and again in 2003, American Rivers, a national nonprofit, recognized the Ipswich River as one of the most threatened or endangered rivers in America, primarily due to severe low flow problems (American Rivers 1997, 2003, IRWA 2003, Zarriello and Reis 2000). Much of the upper half of the River dried up or was reduced to isolated stagnant pools in the summers of 1995, 1997, 1999, 2001, 2002, 2003, and 2005. In 1999, the River experienced record low-flows in May, June, July and August. Major fish kills were documented in 1995, 1997, 1999, 2002, and 2005.

The primary causes of impairments in the Ipswich River watershed are low flow alterations from groundwater withdrawals and runoff from impervious surfaces. This results in a loss of groundwater that supports the baseflow of the river between precipitation events. Low flows have the effect of causing the river to heat more rapidly in the summer. Additional warming in the summer is caused by drought conditions as well as stormwater runoff directly entering the river from paved areas when runoff is typically much warmer than groundwater. Under these conditions, dissolved oxygen levels decrease below what is suitable to aquatic life such as fish and macroinvertebrates that are an important part of the aquatic food web.

Low flows in summer have been linked to ground water withdrawals, particularly in the upper watershed (Zarriello and Reis 2000). Additionally, the diversion of wastewater to treatment plants outside the watershed also significantly reduces flow (Ibid). Many sub-basins in the watershed experience severe flow depletion seasonally due to groundwater withdrawals and significant annual flow depletion due to surface water withdrawals (Weiskel, *et al.* 2009).

Water quality impairments are also caused by flow alterations from dams and road-stream crossings. There are 3 major dams on the mainstem of the river and approximately 70 throughout

the watershed. Dams create pond-like impoundments, impair habitat and block important fish Streams are also segmented to some degree by the roughly 500 road-stream crossings.

Low flow problems have resulted in the loss of flow dependent fish species that would otherwise occur in the Ipswich River (Armstrong et al. 2001). The study identified critical aquatic habitats and recommended minimum flows necessary to preserve those habitats. The Ipswich River Fisheries Restoration Task Group then developed recommendations to restore healthy fisheries to the Ipswich River (2002). These recommendations include maintaining flow over riffle areas, maintaining water to the channel margins and maintain seasonal flow variations near natural levels (Ibid).

Under the Massachusetts Surface Water Quality Standards (MassDEP 1996), most of the freshwater section of the Ipswich River is classified as a Class B water body and warm water fishery, except for public water supplies and certain tributaries (Table 1). The water quality goal for Class B waters is to be "fishable and swimmable" throughout the year. The tidal section of the river located downstream of the Ipswich Mills Dam in Ipswich is classified as a class SA water body. Class SA water bodies are tidal waters intended to be fishable, swimmable, and safe for shell fishing. Table 2 details the water quality standards associated with these classifications.

The Massachusetts Department of Environmental Protection (MassDEP) monitors surface water quality and develops a plan to bring back into compliance those waters that do not meet standards. Under section 303d of the Clean Water Act, states are required to report a list of impaired waters and in the proposed 2014 list; all sections of the Ipswich River were designated as impaired (MassDEP, 2014). A watershed monitoring program on a 5 year rotating schedule is implemented by MassDEP to identify and rank impaired waterbodies. In the 2000 Water Quality Assessment Report for the Ipswich River watershed, 91% of the named river miles throughout the watershed were assessed and 53% of these were impaired for supporting healthy populations of aquatic life (Mass DEP, 2000).

The RiverWatch water quality monitoring program is an effort to provide high quality data on the health of the Ipswich River in order to make informed decisions about water management practices and monitor ongoing restoration efforts.

Our thanks to our volunteers that have monitored on sunny and rainy days, in cold and heat, and high and low river flows. Thank you for your considerable efforts and dedication to the Ipswich River!

Table 1. Massachusetts surface water classifications for the Ipswich River watershed and coastal drainage area (MassDEP, 2007).

| BOUNDARY | MILE POINT | CLASS | OTHER RESTRICTIONS |
|--|---------------|-------|---|
| Ipswich River | | | |
| Source to Salem Beverly Waterway Canal | 41.1 - 16.4 | В | Treated Water Supply, Warm Water, High Quality Water |
| Salem Beverly Waterway Canal to tidal portion | 16.4 - 4.5 | В | Warm Water, High Quality Water |
| Tidal portion and tributaries thereto | 4.5 - 0.0 | SA | Shellfishing (O) |
| Middleton Pond | | | |
| Source to outlet in Middleton and those tributaries thereto | - | A | Public Water Supply |
| Swan Pond | | | |
| Source to outlet in North Reading and those tributaries thereto | - | A | Public Water Supply |
| Mill Pond | | | |
| Source to outlet in Burlington and those tributaries thereto | - | A | Public Water Supply |
| Longham Reservoir | | | |
| Source to outlet in Wenham and those tributaries thereto | - | A | Public Water Supply |
| Wenham Lake | | | |
| Source to outlet in Wenham and those tributaries thereto | - | A | Public Water Supply |
| Putnamville Reservoir | | | |
| Source to outlet in Danvers and those tributaries thereto | - | A | Public Water Supply |
| Suntaug Lake | | | |
| Source to outlet in Lynn and Peabody and those tributaries thereto | - | A | Public Water Supply |
| Winona Pond | | | |
| Pond to outlet in Peabody and those tributaries thereto | - | A | Public Water Supply |
| Unnamed Reservoir (Emerson Broo | k Reservoir) | | |
| Reservoir to outlet in Middleton and those tributaries thereto | - | A | Public Water Supply |

Table 2. Massachusetts Department of Environmental Protection water quality standards (2007).

| · · · | | |
|---|---|---|
| | Class B Standards | Class SA Standards |
| AQUATIC LIFE Dissolved Oxygen | 5.0 mg/L * | 6.0 mg/L |
| Temperature | 83° F Max ** (28.3° C) | 85 F (29.4° C) Max, 80 F Average |
| pН | 6.5 - 8.3 | 6.5 - 8.5 |
| PRIMARY CONTACT RECREATION Fecal Coliform | 200 / 100 mL geo. mean 10% <= 400 / 100 mL | 200 / 100 mL geo. mean 10% <= 400 / 100 mL |
| SECONDARY CONTACT RECREATION Fecal Coliform | 1000 / 100 mL geo. mean 10% <= 2000 / 100 mL | $1000 / 100 \; \text{mL}$ geo. mean $10\% <= 2000 / 100 \; \text{mL}$ |
| SHELLFISHERY Fecal Coliform | Not applicable | 14 / 100 mL geo. mean 10% <= 43 / 100 mL |
| AESTHETICS Taste and Odor | None that are objectionable | None other than natural |
| ** Warm water fishery | | |

^{**} Warm water fishery.

1314 CMR 4.05 (3) (b)1.b. states that Dissolved Oxygen "levels shall not be lowered below...60% of saturation in warm water fisheries due to a discharge." This report will therefore assume 60% of saturation to be the Class B standard.

In 2008, the State eliminated standards pertaining to DO% saturation. Values in this report are based on the previous standard of a minimum of 60% DO saturation and presented for comparison with previous years.

2314 CMR 4.05 (4)(a)1.b.states that Dissolved Oxygen "levels shall not be lowered below 75% of saturation due to a discharge." This report will therefore assume 75% of saturation to be the Class SA standard.

2.1 Program Description and Monitoring Methods

Monthly Water Quality Testing

As stated earlier, IRWA has conducted informal monitoring from 1988-1996. The RiverWatch program took its current form in 1997 and has been continuously monitoring the Ipswich River Watershed since this time. In order to best use our resources to gain an accurate picture of the Ipswich River, 10 tributary sites and 22 sites along the mainstem of the River from Wilmington to Ipswich, have been identified for monitoring once a month from March through December (table 3). Both Fish Brook at Brookview Farm Rd. (FB-BV) and Greenwood Creek (GC) were discontinued in 2001.

Volunteer monitors are responsible for monthly monitoring which takes place in the morning of the last Sunday of each month from March through December unless the date conflicts with a holiday, in which case, the previous or next Sunday will be chosen. All samples are collected between 8 am and 12:30 pm, except for the tidal locations, which are sampled within 1 hour of low tide closest to the 8 am to 12:30 pm time span. Sampling in the morning is extremely important because the lowest dissolved oxygen values are generally observed in the early morning. This is desirable, because low values have the most potential to affect the organisms living in the Ipswich River. As of the spring of 2006, sampling in January and February became optional. Historically, volunteers sampled during these months, but the River was often frozen and the data collected during these months was generally not used in management decisions.

Volunteers record information on weather, rain in the last 48 hours and river status (frozen or dry). Monitors then collect a grab sample using a bucket. While water is contained in the sampling bucket, observations of color, clarity and odor are made. Color is recorded as a range of pre-determined colors from Clear to Dark Tea. Clarity is recorded as the amount of turbidity in the water from a scale ranging from clear to highly turbid.

Water temperature is measured followed by a test for dissolved oxygen. Water Temperature is measured with H-B Enviro-Safe® Thermometers. Monitors are asked to round to the nearest 0.5 degrees Celsius.

Dissolved Oxygen (DO) is measured with a LaMotte Modified Winkler Method Test Kit. One drop of fluid from the direct reading titrator in the kit is approximately 0.4 mg/L. Thus, accuracy from the titrator is +/- 0.2 mg/L of dissolved oxygen. Field audits are conducted once per year comparing results from DO kits with results from other test kits or a dissolved oxygen meter, obtained by the trainer, with a goal of all sites being within 1mg/L of measure DO concentration. In addition, duplicate DO samples were taken at each site at least once during the monitoring year.

For DO, a percent saturation value is also calculated. This is a percentage of the DO measured in the water relative to the maximum DO water could theoretically hold at the testing water temperature (and elevation).

Velocity is measured by dividing the average of three times that it takes an orange peel to travel a known distance (often the width of a bridge). If times are disparate, another three readings are taken. Velocity measurements are multiplied by a correction factor of 0.85.

Depth is measured at a consistent location on the bridge with a weight attached to a decimal measuring tape. Cross-sections are taken at monitoring sites located at selected bridges twice each year (April and September). Monitors try to take up to 20 measurements across the channel at one or two foot increments. On the cross section data sheet, volunteers indicate at what location they measure depth each month. A cross-section profile is plotted and an approximate flow value can be calculated by adding the product of average velocity by each cross-sectional area.

Conductivity is measured at selected sites as an indicator of human impact from sources such as stormwater runoff. Ions from sources such as road salts and leaking septic systems increase conductivity which can negatively impact aquatic life. All nine tributary sites are monitored for conductivity since these may be expected to vary more than along the mainstem of the river where five sites are monitored to detect variations. This is done using an Oakton Eco Testr EC Low or Oakton ECTestr Low conductivity meter. The meter is first rinsed with deionized or distilled water. The meter is calibrated using 447 μ Siemens/cm conductivity standard solution. The meter is rinsed again and placed in the sampling bucket to record the conductivity value.

As stated previously, data collected will be reported to IRWA members, state agencies, interested organizations, and conservation commissions through reports and presentations on the collected data. Atypical data will be reported to the appropriate agencies. Atypical data include dissolved oxygen data that vary significantly from adjacent sites over one or more months. Extended periods of no flow or extremely low dissolved oxygen (less than 2 mg/L) are also considered extremely important and will be presented to state agencies. (When dissolved oxygen levels fall below 2 mg/L the health of fish and other aquatic organisms can be severely impacted.)

For data to be reported to state agencies, a Quality Assurance Project Plan (QUAPP) is maintained with MassDEP and most recently updated for the period 2013-2015. The QUAPP requires all new and returning monitors to receive annual training, and an annual site audit of each volunteer. Prior to monitoring, new monitors receive a walk-through of the monitoring manual and hands-on training at a monitoring site. All new and returning monitors must attend an annual training that consists of an overview of the program and procedures followed by a collection and analysis of temperature, dissolved oxygen and conductivity samples for comparison with readings obtained by the Monitoring Project Coordinator. Records of data generated during this training as well as attendance records are retained by IRWA.

During the year, each site is audited by the Monitoring Project Coordinator. This consists of the observation of the volunteer by the auditor. Any errors in procedure are recorded on the project audit sheet and problems discussed and resolved with the volunteers.

Table 3. Monitoring site information. Note that macroinvertebrate Sampling is discussed in a separate report.

| Site | Location | Town | Latitude | Longitude | Date Start | Dissolved Overson | Tomporaturo | Donth | Volocity | Cross Soction | Conductivity | Staff Caugas | Macroinvertebrates |
|---------|---|---------------|-----------|------------|------------------|-------------------|-------------|----------|----------|---------------|--------------|--------------|----------------------|
| MMB | Maple Meadow Brook, Wildwood Street | Wilmington | 42.552842 | -71.156567 | Aug-97 | | √ √ | ν √ | √ | Cross Section | ✓ | Starr Gauges | iviacionivertebrates |
| LB | Lubbers Brook, Glen Rd. | Wilmington | 42.552642 | -71.130307 | Aug-97 Aug-97 | | <u> </u> | √ | √ | | | | |
| IP00 | Woburn St. | Wilmington | 42.553750 | -71.182/92 | Jan-97 | · | → | √ | √ | | → | | |
| IP00.5 | Reading Town Forest | Reading | 42.554464 | -71.110633 | Nov-97 | ✓ | → | √ | √ | | <u> </u> | | |
| IP00.5 | Mill St. | | 42.554464 | -71.107633 | | √ | ∨ | ∨ | <u> </u> | | • | | |
| | | Reading | | | Jan-97 | √ | ✓ | ∨ | ✓ | √ | | | |
| IP02 | Route 28 | Reading | 42.564583 | -71.107633 | Jan-97 | · | ✓ | | ✓ | · · | ✓ | | |
| MB-62 | Martins Brook, Rt. 62 | Wilmington | 42.579774 | -71.138944 | Jan-11 | | | ✓ | | | | | , |
| MB-PS | Martin's Brook, Park Street | North Reading | 42.571475 | -71.101233 | Mar-99 | ✓ | ✓ | ✓ | ✓ | | ✓ | ✓ | √ |
| IP2.7 | Parish Park | North Reading | 42.571783 | -71.094967 | Jan-99 | | | | | | | | ✓ |
| IP3.5 | Haverhill St. | North Reading | 42.572425 | -71.080336 | Jun-12 | | | | | | | ✓ | |
| IP03 | Central St. | North Reading | 42.570047 | -71.029386 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | | | |
| IP04 | Washington St. (Route 62) | North Reading | 42.576553 | -71.069583 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | | |
| IP06 | South Middleton Gage | Middleton | 42.570047 | -71.029236 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | | | ✓ |
| IP08 | Log Bridge Road | Middleton | 42.577892 | -70.996964 | Mar-99 | ✓ | ✓ | ✓ | ✓ | | | | |
| IP10 | Maple St. (Route 62) | Middleton | 42.595131 | -70.997014 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | | |
| BB | Peabody St. | Middleton | 42.620317 | -71.020308 | Oct-14 | | | | | | | | ✓ |
| IP11 | Peabody St. | Middleton | 42.616442 | -70.996964 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | | | |
| IP12 | Thunder Bridge (East St.) | Middleton | 42.619575 | -70.988239 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | | | |
| FB-BV* | Fish Brook, Brookview Farm Rd. | Boxford | 42.662897 | -71.030072 | Jan-97 | | | | | | | | |
| FB-MR | Fish Brook, Mill St. | Boxford | 42.655261 | -70.999325 | Nov-14 | | | | | | | ✓ | |
| FB-MI | Fish Brook, Middleton Rd. | Boxford | 42.658294 | -71.143658 | Mar-99 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | | |
| FB-LL | Fish Brook, Lockwood Ln. | Boxford | 42.645247 | -70.989189 | Oct-13 | | | | | | | | ✓ |
| FB-WA | Fish Brook, Washington St. | Boxford | 42.630628 | -70.973783 | Mar-99 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | | |
| IP13 | Rowley Bridge Road | Topsfield | 42.627017 | -70.966953 | Jan-97 | ✓ | ✓ | ✓ | ✓ | √ | | | |
| IP14 | Salem Road | Topsfield | 42.625722 | -70.949758 | Jan-97 | ✓ | ✓ | ✓ | ✓ | ✓ | ✓ | | |
| IP16 | IRWS - Boat Launch | Topsfield | 42.627197 | -70.917922 | Jan-97 | ✓ | ✓ | ✓ | | | | | |
| HB . | Howlett Brook, Topsfield Rd | Ipswich | 42.654839 | -70.917539 | Mar-99 | ✓ | √ | ✓ | ✓ | | √ | | ✓ |
| IP18 | Asbury Road | Topsfield | 42.653761 | -70.911933 | Jan-97 | ✓ | ✓ | | | | | | |
| GB | Gravelly Brook, Willowdale State Forest | Ipswich | 42.661817 | -70.903883 | Jun-11 | ✓ | ✓ | ✓ | ✓ | √ | √ | | ✓ |
| IP19A | 100' Above Willowdale Dam | Ipswich | 42.659917 | -70.894683 | Mar-10 | ✓ | ✓ | | | | | | |
| IP19 | Below Willowdale Dam | Ipswich | 42.659864 | -70.894367 | Jan-97 | ✓ | ✓ | √ | √ | | | | |
| IP20 | Winthrop Street | Ipswich | 42.658706 | -70.890539 | Jan-97 | ✓ | ✓ | 1 | √ | | | | √ |
| IP22 | Mill Road | Ipswich | 42.658372 | -70.861939 | Jan-97 | √ | √ | 1 | √ | | | | |
| IP24 | Sylvania Dam | lpswich | 42.677539 | -70.837686 | Jan-97 | √ | √ | √ | √ | | | | |
| MR-1A | Miles River, Rt. 1A | lpswich | 42.657800 | -70.843431 | Mar-99 | · · | · · | · / | · · | | √ | | |
| ER-1A | Egypt River, Rt. 1A | Ipswich | 42.698179 | -70.869172 | Mar-11 | · · | → | <i>'</i> | | | | | |
| GC* | Greenwood Creek | Ipswich | 42.692494 | -70.839800 | Sep-97 | • | • | , | | | | | |
| IP25 | Green Street | Ipswich | 42.692494 | -70.839800 | Sep-97 Jan-97 | √ | ✓ | 1 | ✓ | | | | |
| IP25 | Town Landing | 1. | | | | √ | ✓ | • | · · | | | | |
| | Ŭ | lpswich | 42.683522 | -70.830467 | Jan-97 | · · | | | | | | | |
| "Discon | tinued in 2001 | | | | | | | | | | | | |

Streamflow Monitoring

Having adequate amounts of flowing water is essential for the health of rivers and streams. The Ipswich River has a history of flow alterations from water withdrawals, particularly in the upper watershed, so measuring streamflow is important to understanding low-flow impacts.

Two real-time streamflow gages are maintained by USGS on the Ipswich River in <u>South Middleton</u> and <u>Ipswich</u> that transmit real-time discharge data. These gages have recorded flow levels since the 1930's, as both a historical record of the river and vital source of real-time information needed to manage municipal water supplies. However, many sections of the river and streams in the watershed are not gaged.

Beginning in 2012, additional streamflow gages have been added to further document streamflow patterns. The Massachusetts Division of Ecological Restoration (MassDER), River Instream Flow Stewards (RIFLS) program enables local groups to monitor streamflow to investigate any signs of flow alteration, with the goal of restoring more natural flow patterns.

Three flow monitoring sites have been established based on the need for additional monitoring flow alterations in these areas. Two sites are located in North Reading; Martins Brook at Park St. (MB-PS) and a new site on the Ipswich River at Haverhill St., designated IP3.5 (table 3). A new staff gage was installed on the bridge abutment at IP3.5 in June 2012, while an existing staff gage that was part of a flow monitoring station maintained by USGS from 2007-2009 is now being used as the RIFLS gage on Martins Brook. A third gage was installed on Fish Brook in Boxford in November 2014. Volunteers read staff gages at these sites on a regular basis and enter data to the RIFLS website (www.rifls.com) where it is converted to a streamflow value in cubic feet per second (cfs) from rating curves maintained by the RIFLS staff with MassDER. Water level data loggers were generously donated by the Nor'East Chapter of Trout Unlimited and installed at all the 3 RIFLS site locations and activated beginning in June 2014. These loggers will collect frequent data that will allow for analysis as detailed as the USGS gages.

Analysis is conducted by downloading data from the RIFLS and USGS websites. Individual gage data are compared by converting mean daily streamflow values from cfs to cubic feet per second per square mile (cfsm). The drainage area values needed for this conversion are obtained from either the USGS or RIFLS websites for each gage. Daily discharge values in cfsm are plotted together and compared. When normalized for area, flows at the RIFLS and USGS gages should be similar. Differences may indicate a flow alteration such as from groundwater pumping. Groundwater pumping records can be used to identify the source, which is the focus of ongoing work.

Section 3: Monthly Water Quality Testing

3.1 Monthly RiverWatch Monitoring Results by Parameter

Temperature

In 2014, all samples met the Class B standard or Class SA standard for maximum water temperature. The Class B standard is a maximum of 28.5° Celsius (83°F); the Class SA standard is a maximum of 29.4° Celsius (85°F), and applies to the tidal sites of IP25 and IP26.

Temperature is an important measure of water quality, as temperatures higher than the natural observed range can reduce the amount of dissolved oxygen that the water can hold (more on dissolved oxygen in the next section). This can create a stressful environment for aquatic organisms. For example, some fish, like brook trout, cannot survive in warm water.

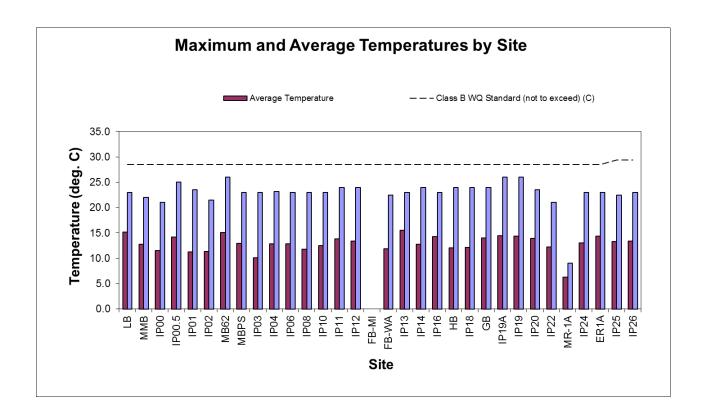
Annual Statistics

Table 3 is a summary of annual statistics for temperature. Temperature has exceeded the state standard only 5 times since 1997. This does not reflect the times the river has dried up and monitoring could not take place. Figure 4 is a comparison of average annual and maximum water temperature for 2014.

Table 3: Annual temperature statistics for all sites.

| | # Samples (March- | | | Annual Average (°C) | Summer Average | # Samples Outside Class | % Violations (% of samples |
|---------------|-------------------|--------------|--------------|---------------------|--------------------|-------------------------|----------------------------|
| Year | December) | Minimum (°C) | Maximum (°C) | (March-December) | (°C) (June-August) | B, Class SA Standard | not meeting standard) |
| 1997 | 172 | -4 | 31.0 | 11.6 | 21.0 | 2 | 1.2% |
| 1998 | 227 | 1 | 26.0 | 14.5 | 21.2 | 0 | 0.0% |
| 1999 | 257 | 0 | 28.0 | 13.7 | 22.9 | 0 | 0.0% |
| 2000 | 247 | 0 | 25.0 | 13.1 | 20.5 | 0 | 0.0% |
| 2001 | 236 | -3 | 25.3 | 12.1 | 20.4 | 0 | 0.0% |
| 2002 | 241 | 0 | 25.5 | 11.7 | 20.2 | 0 | 0.0% |
| 2003 | 226 | 0 | 29.0 | 12.8 | 21.6 | 1 | 0.4% |
| 2004 | 234 | -2 | 25.0 | 12.2 | 20.2 | 0 | 0.0% |
| 2005 | 237 | -2.5 | 34.0 | 12.1 | 22.0 | 2 | 0.8% |
| 2006 | 212 | 0 | 28.0 | 13.1 | 21.3 | 0 | 0.0% |
| 2007 | 213 | 0 | 26.0 | 13.8 | 21.7 | 0 | 0.0% |
| 2008 | 209 | -1 | 29.0 | 13.1 | 21.2 | 1 | 0.5% |
| 2009 | 202 | 0 | 24.0 | 14.2 | 19.2 | 0 | 0.0% |
| 2010 | 217 | 0 | 27.5 | 14.7 | 22.5 | 0 | 0.0% |
| 2011 | 224 | 0.5 | 26.0 | 12.8 | 20.6 | 0 | 0.0% |
| 2012 | 266 | 0 | 30.0 | 14.2 | 22.2 | 1 | 0.4% |
| 2013 | 234 | 1 | 26.0 | 14.7 | 21.8 | 0 | 0.0% |
| 2014 | 274 | 0.5 | 26.0 | 13.3 | 21.7 | 0 | 0.0% |
| Entire Record | 4128 | -0.5 | 27.3 | 13.2 | 21.2 | 7.0 | 0.2% |

Figure 4: Maximum and Average Water Temperatures, by Site, 2014. The dashed line indicates the maximum temperature for class B (28.5°C) and Class SA waters (29.4°C).



General Findings

Water temperature readings met state standards throughout 2014 across the watershed (i.e., temperatures remained below the state standard maximum temperature). It is important to note that recorded temperatures are conservative, as temperatures are not recorded when there is no water present in the river during extreme low flows. Also, monitoring is conducted in the morning, and may not represent the highest temperatures that occur in the course of that day or month.

Dissolved Oxygen

The amount of dissolved oxygen (DO) in water depends on numerous factors, including the temperature of the water and the gas exchange across the air-water interface. DO can increase when water is at lower temperatures and in areas where there is turbulence in the water (e.g., riffles or rapids). Other primary factors affecting DO include oxygen production through photosynthesis and depletion through respiration and other oxygen-demanding processes. DO changes on a diurnal basis as well as seasonally, and is affected by cloud cover and other weather conditions. The most critical time for organisms is in the early morning hours on hot summer days when water temperatures are high, flows are low and photosynthesis has ceased producing oxygen since sunset. The interactions of factors affecting DO in

the natural environment are quite complex, and a full exploration of this topic is beyond the scope of this report, but warrants further investigation.

Sampling was conducted during morning hours because DO is typically lowest at or just after dawn, so morning sampling is likely to capture relatively low DO. Therefore the values observed generally represent a more stressed condition than if the values were mid-day or later.

For dissolved oxygen concentration, the Class B standard requires a minimum of 5.0 mg/L; the Class SA standard is a minimum of 6.0 mg/L DO, and applies to the tidal sites of IP25 and IP26. For dissolved oxygen percent of saturation, 60% is considered the minimum for good water quality in class B waters and 75% in class SA waters. The state of Massachusetts no longer uses the standard for percent of saturation; however, we continue to monitor according to this figure.

Table 4 presents annual statistics for DO concentration and percent saturation for all sites monitored. The number of samples for percent saturation can differ from concentration if either a concentration or water temperature value is missing since it is calculated from both.

Annual Statistics

Table 4: Annual statistics for dissolved oxygen concentration (A) and percent of saturation (B) for all sites.

Note:In 2008, the State eliminated standards pertaining to DO% saturation. Numbers are presented for comparison with previous years and are based on the previous standard of 60% saturation for class B and 75% saturation for class SA sites.

Table 4 A. Dissolved Oxygen Concentration (mg/L)

| | 2188017 | • • | | | | | % Violations (% of |
|---------------|-------------------|---------|---------|-----------------------|-----------------------|-------------------------|---------------------|
| | # Samples (March- | Minimum | Maximum | Annual Average (mg/L) | Summer Average (mg/L) | # Samples Outside Class | samples not meeting |
| Year | December) | (mg/L) | (mg/L) | (March-December) | (June-August) | B, Class SA Standard | standard) |
| 1997 | 100 | 1.0 | 14.4 | 7.6 | 6.1 | 30 | 30% |
| 1998 | 230 | 0.0 | 12.2 | 6.3 | 4.1 | 78 | 34% |
| 1999 | 262 | 0.4 | 14.8 | 7.3 | 5.0 | 65 | 25% |
| 2000 | 264 | 1.0 | 14.0 | 7.1 | 5.2 | 56 | 21% |
| 2001 | 240 | 0.2 | 14.0 | 6.9 | 4.6 | 73 | 30% |
| 2002 | 239 | 0.2 | 12.4 | 7.1 | 5.3 | 57 | 24% |
| 2003 | 225 | 0.1 | 12.4 | 6.5 | 3.9 | 75 | 33% |
| 2004 | 240 | 0.0 | 12.4 | 6.6 | 4.3 | 61 | 25% |
| 2005 | 240 | 0.6 | 13.2 | 6.8 | 4.5 | 62 | 26% |
| 2006 | 213 | 0.2 | 13.0 | 6.4 | 4.1 | 74 | 35% |
| 2007 | 216 | 0.6 | 16.2 | 6.3 | 4.9 | 68 | 31% |
| 2008 | 207 | 0.6 | 13.0 | 6.6 | 4.0 | 71 | 34% |
| 2009 | 203 | 0.8 | 12.7 | 6.1 | 4.5 | 72 | 35% |
| 2010 | 219 | 0.0 | 12.6 | 6.3 | 4.5 | 69 | 32% |
| 2011 | 205 | 0.6 | 12.6 | 7.2 | 4.6 | 56 | 27% |
| 2012 | 270 | 0.5 | 14.0 | 6.2 | 4.1 | 86 | 32% |
| 2013 | 239 | 0.1 | 13.4 | 6.2 | 4.0 | 82 | 34% |
| 2014 | 277 | 0.4 | 12.6 | 6.4 | 4.4 | 93 | 34% |
| Entire Record | 4089 | 0.4 | 13.3 | 6.7 | 4.5 | 1228.0 | 30% |

Table 4 B. Dissolved Oxygen Percent of Saturation

| | | | | | | # Samples Outside | % Violations (% of |
|---------------|-------------------|---------|---------|------------------|----------------|-------------------|--------------------|
| | # Samples (March- | | | Annual Average | Summer Average | Class B, Class SA | samples not |
| Year | December) | Minimum | Maximum | (March-December) | (June-August) | Standard | meeting standard)* |
| 1997 | 89 | 7.8 | 122.6 | 66.0 | 67.6 | 27 | 30% |
| 1998 | 224 | 0.0 | 101.2 | 59.2 | 45.7 | 109 | 49% |
| 1999 | 249 | 4.4 | 101.7 | 67.7 | 58.1 | 84 | 34% |
| 2000 | 239 | 11.7 | 112.9 | 64.1 | 56.7 | 98 | 41% |
| 2001 | 214 | 2.2 | 105.5 | 61.1 | 51.8 | 103 | 48% |
| 2002 | 231 | 2.1 | 119.7 | 63.8 | 58.6 | 96 | 42% |
| 2003 | 217 | 0.7 | 99.2 | 58.9 | 43.7 | 105 | 48% |
| 2004 | 229 | 0.0 | 97.4 | 59.1 | 47.4 | 108 | 47% |
| 2005 | 227 | 6.7 | 115.9 | 59.9 | 50.9 | 109 | 48% |
| 2006 | 209 | 2.4 | 117.9 | 58.2 | 45.4 | 107 | 51% |
| 2007 | 207 | 6.2 | 123.6 | 59.0 | 54.6 | 112 | 54% |
| 2008 | 197 | 6.5 | 104.0 | 58.7 | 45.1 | 96 | 49% |
| 2009 | 199 | 9.1 | 112.5 | 58.1 | 48.3 | 104 | 52% |
| 2010 | 216 | 0.0 | 94.6 | 59.0 | 51.8 | 103 | 48% |
| 2011 | 203 | 6.9 | 115.5 | 64.9 | 51.3 | 84 | 41% |
| 2012 | 262 | 5.7 | 98.5 | 57.7 | 46.1 | 144 | 55% |
| 2013 | 234 | 1.2 | 110.0 | 58.5 | 45.7 | 116 | 50% |
| 2014 | 274 | 0.0 | 100.4 | 57.9 | 49.9 | 144 | 53% |
| Entire Record | 3920 | 4.1 | 108.5 | 60.7 | 51.1 | 1849 | 47% |

In 2014, 34% of all samples taken by volunteers did not meet the combined state standards of 5 mg/L for class B and 6mg/L for class SA waters (93 of 277 samples). When calculating percent saturation of dissolved oxygen, 53% of these same samples fell below the combined standards of 60% saturation for class B and 75% saturation for class SA waters.

Site Statistics

Low DO conditions have been widespread and frequent during the past 18 years of monitoring. In 2014:

- Summer averages for 20 sites (out of 32) were less than 5.0 mg/L DO concentration. Nine sites had summer DO averages below 3.0 mg/L (figure 6).
- Annual averages for 5 (out of 32) sites were less than 5.0 mg/L DO concentration.
- Twenty seven sites out of 32 had a minimum DO concentration below 5.0 mg/L DO. Only 5 sites had minimum values above 5.0 mg/L.
- Values at one of the tidally influenced sites (IP25) fell below 6mg/L on three instances.
- 30% of the 244 samples for dissolved oxygen were below the standard for concentration (5 mg/L).

Figure 6 shows average and minimum dissolved oxygen concentration values for all sites in 2014, while figure 7 shows the distribution of sites with low dissolved oxygen relative to river health.

The fact that DO levels were very low consistently over the past decade represents a significant impaired condition on the river, and indicates that many aquatic organisms are under high stress conditions. Many organisms may not likely survive during most summers.

Figure 6. Average annual and minimum dissolved oxygen concentration (mg/L) for all sites monitored in 2014. The dashed line indicates the minimum standard for class B (5.0 mg/L) and class SA waters (6.0 mg/L)

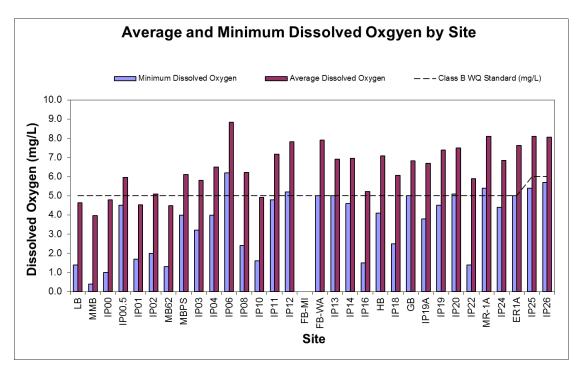
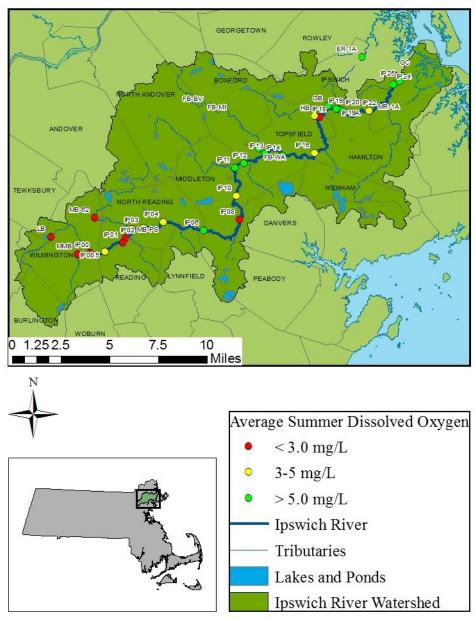


Figure 7. Average summer dissolved oxygen levels for 2012 and relative river health.

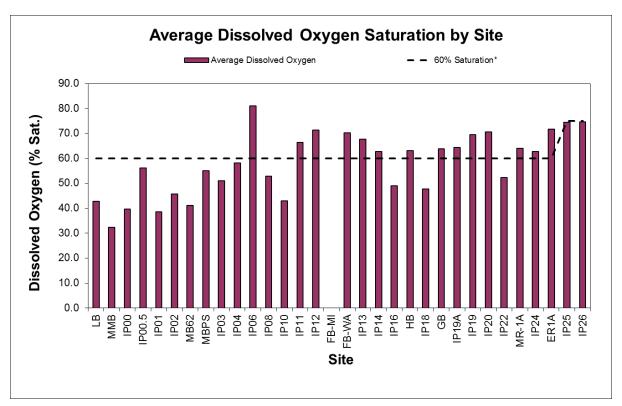
RiverWatch Monitoring Sites



- > 5mg/L (Class B), > 6mg/L (Class SA): Supports aquatic organisms.
- 3-5 mg/L: organisms may become stressed.
- < 3mg/L: Mobile organisms will move to areas of higher DO and immobile organisms may die.</p>
 - <0.5 mg/L Cannot support most aquatic life.

Dissolved oxygen, percent of saturation is defined as the amount of oxygen that can be absorbed by water at a given temperature. Colder water can absorb more oxygen than warmer water. The Commonwealth of Massachusetts discontinued use of a water quality standard for dissolved oxygen, percent of saturation in 2008, but the data are presented here for comparison with dissolved oxygen concentration (mg/L) measurements and for comparison with previous years. The previously used standard of 60% (class B) and 75% (class SA) saturation can be used to confirm water oxygen depletion, particularly in the upper watershed. Most sites in the upper watershed did not achieve 60% saturation over the course of the year and especially in summer months when water temperatures are highest. Site statistics for dissolved oxygen, percent saturation are presented in table 4 and figure 8.

Figure 8: Average Dissolved Oxygen Percent Saturation Statistics for 2014.* The dashed line represents the minimum standard for class B waters (60%) and class SA waters (75%).



^{*}In 2008, the state discontinued use of the 60% saturation standard for dissolved oxygen percent saturation. Values are presented here for comparison with previous years.

General Findings

Dissolved oxygen (DO) is usually lowest at or shortly after dawn, and then increases during daylight hours. Sampling was conducted during morning hours, likely capturing lower DO than what occurs in the afternoon, and therefore the values observed represent the lower end of the daily DO fluctuation.

Frequent and prolonged low DO conditions represent a serious threat to aquatic organisms that are dependent on the river for survival. State standards represent a minimum condition that is protective of the health of aquatic organisms and the Ipswich River repeatedly and for extended periods of time does not meet those minimum standards. Fish kills were observed in 1995, 1997, 1999, 2002, and 2005. Under natural conditions, DO varies considerably daily and seasonally, as well as in response to weather conditions and numerous other factors, so conclusively stating the causes of the extremely low DO documented on the Ipswich River is beyond the scope of this report. It might be expected that DO levels in the Ipswich River tend towards the lower end of that 5-10 mg/L healthy DO concentration range because of the relatively low gradient of the river and the presence of numerous wetlands and forest that contribute organic matter (like leaves) to the water. For example, sites IP08 and IP18 are both located downstream of wetlands. Both sites exhibit average summer DO levels lower than other surrounding sites (figures 6 and 7). However, the Ipswich River experiences DO levels that fall consistently lower than this natural range, and consistently lower than state standards for a healthy river.

A statistical investigation into the causes of low DO was conducted by IRWA in 2002, and indicated that variables most linked with DO levels are water temperature, river kilometer (how far upstream the site is), depth, and the previous 28-day rainfall amount (IRWA, 2002). While this study provides a first step towards better understanding of variation in DO in the Ipswich River, there remain a number of unanswered questions warranting further study. For example: what is the role of these variables and their interactions on DO levels; what are the causes of the observed changes in these variables; what is the extent and health of wetlands adjacent to the river; and, how can management actions and behavioral changes alleviate low DO levels in the river?

Depth, Velocity and Streamflow

Depth and velocity are measured as rough indicators of channel coverage and flow at individual sites. Because depth is measured from the middle of the channel at most sites, generally it is an optimistic indicator of depth across the channel, since drying will typically occur first at the channel margins. There are, however, occasions when flow is too high to accurately measure depth (or velocity), such as during the flooding event in May of 2006 and March 2010. Conversely, velocity is a conservative indicator, since volunteers insert the floatable object only where there is noticeable current. Immeasurable velocities cannot be quantified.

Water velocity is measured as an indicator of the amount of flow in the river. Monitors record the time it takes a floating object such as an orange peel to travel a known distance, usually the width of the bridge spanning the river or between two points along the bank. Only sites with a bridge or where it is convenient to do so will measure velocity. Water velocity is typically lowest in the upper watershed where there is a low gradient to the river and tributaries and surrounding wetlands (figure 10). Sites IP01 and IP03 are located at bridges where the channel width narrows, increasing water velocity during spring runoff events beyond what would be expected naturally.

Flow is an obvious and important measure of river health. Observations of a dry riverbed or very low flow associated with very small amounts of water in the river are indicative of a serious impairment. Unfortunately, numerous episodes of little or no flow have been documented for the Ipswich River.

The United States Geological Survey (USGS) maintains two real-time streamflow gauges on the Ipswich River. One is located near Boston St. in South Middleton and the other is located near Topsfield Rd. and Winthrop St. in Ipswich. Water depth or stage height is recorded and compared to a rating curve of flow measurements taken over time at high and low water levels. The result is a flow volume measured in cubic feet per second (cfs). The South Middleton and Ipswich gauges have been recording streamflow data since the 1930's.

These gauges have recorded regular episodes of extended extreme low flow events over the past 18 years. "Extreme low flow" is defined by the USGS as discharge levels below a minimum summer "ecological protection flow" (Horsley and Witten 2002). This "ecological protection flow" is the flow that "provides adequate habitat for the protection of fisheries" (Ibid). The summer ecological protection flow for the Ipswich River is 0.42 cubic feet per second per square mile (cfsm).

Summer low flows at the Ipswich gauge are defined as flows lower than 52.5 cfs (calculated as 0.42 cfsm multiplied by the drainage area of 125 square miles). Summer low flows at the South Middleton gauge are defined as flows falling below 18.6 cfs (calculated as above, with a drainage area of 44.5 square miles).

Percent of summer days (June-September) were compared for all flow monitoring gages, including the RIFLS gages. Daily average flows recorded by data loggers at the RIFLS gages allowed these sites to be included in the low flow analysis. In 2014, the percent of summer days experiencing low flows was an average of approximately 60% among all flow gage sites (figure 12). Low flows appear to be decreasing in frequency over time and more observations and analysis will be needed to verify this.

Site Statistics

In 2014, as in previous years, most sites recorded average water depths that were highest during the winter and lowest during the summer (table 5, figure 10).

Table 5. Annual statistics for depth and velocity for all sites.

| | | | | | Annual | Summer |
|--------------------|---------------|---------|---------|---------|-----------|---------|
| | | Number | | | Average | Average |
| Water Quality | | of | | | (March- | (June- |
| Parameter | Year | Samples | Minimum | Maximim | December) | August) |
| | 1997 | 158 | 0.0 | 10.2 | 1.1 | 0.8 |
| | 1998 | 207 | 0.0 | 6.0 | 0.1 | 0.1 |
| | 1999 | 253 | 0.0 | 7.3 | 1.1 | 0.3 |
| | 2000 | 232 | 0.0 | 6.1 | 1.6 | 1.2 |
| | 2001 | 190 | 0.0 | 16.0 | 1.4 | 1.3 |
| | 2002 | 182 | 0.1 | 54.5 | 1.8 | 1.5 |
| | 2003 | 183 | 0.0 | 5.1 | 1.6 | 1.3 |
| (pe) | 2004 | 210 | 0.0 | 25.3 | 1.7 | 2.1 |
| t./s | 2005 | 209 | 0.0 | 23.9 | 1.0 | 0.3 |
| Velocity (ft./sec) | 2006 | 185 | 0.1 | 9.8 | 1.7 | 1.5 |
| ocit | 2007 | 150 | 0.1 | 8.3 | 1.5 | 0.8 |
| Vel | 2008 | 172 | 0.0 | 16.6 | 1.6 | 2.0 |
| | 2009 | 162 | 0.0 | 21.7 | 1.6 | 1.6 |
| | 2010 | 133 | 0.1 | 35.1 | 1.5 | 0.6 |
| | 2011 | 173 | 0.0 | 5.9 | 1.7 | 1.2 |
| | 2012 | 174 | 0.1 | 4.4 | 1.0 | 0.8 |
| | 2013 | 140 | 0.0 | 5.0 | 1.1 | 1.2 |
| | 2014 | 159 | 0.1 | 7.2 | 1.6 | 1.0 |
| | Entire Record | 3272 | 0.0 | 14.9 | 1.4 | 1.1 |
| | 1997 | 141 | 0.0 | 10.0 | 3.1 | 2.6 |
| | 1998 | 221 | 0.0 | 8.8 | 2.7 | 2.8 |
| | 1999 | 248 | 0.0 | 8.0 | 2.3 | 1.6 |
| | 2000 | 244 | 0.0 | 11.3 | 2.9 | 2.4 |
| | 2001 | 219 | 0.0 | 22.0 | 2.6 | 2.1 |
| | 2002 | 224 | 0.0 | 9.2 | 2.7 | 1.8 |
| | 2003 | 198 | 0.0 | 9.3 | 3.1 | 2.5 |
| | 2004 | 209 | | 10.0 | 3.3 | 3.1 |
| (ft.) | 2005 | 200 | 0.0 | 8.5 | 3.0 | 2.2 |
| Depth (ft | 2006 | 192 | 0.4 | 10.7 | 3.6 | 3.1 |
| Dep | 2007 | 189 | 0.1 | 10.6 | 3.3 | 2.5 |
| | 2008 | 192 | 0.4 | 9.7 | 3.4 | 3.4 |
| | 2009 | 177 | 0.5 | 10.7 | 3.3 | 3.5 |
| | 2010 | 186 | 0.1 | 9.3 | 2.8 | 2.1 |
| | 2011 | 204 | 0.2 | 8.5 | 3.4 | 2.7 |
| | 2012 | 237 | 0.2 | 6.4 | 2.4 | 2.1 |
| | 2013 | 190 | 0.0 | 18.6 | 3.0 | 2.7 |
| | 2014 | 211 | 0.1 | 15.4 | 3.1 | 2.1 |
| | Entire Record | 3682 | 0.1 | 10.9 | 3.0 | 2.5 |

Figure 10: Comparison of average annual, summer and winter water velocity by site.

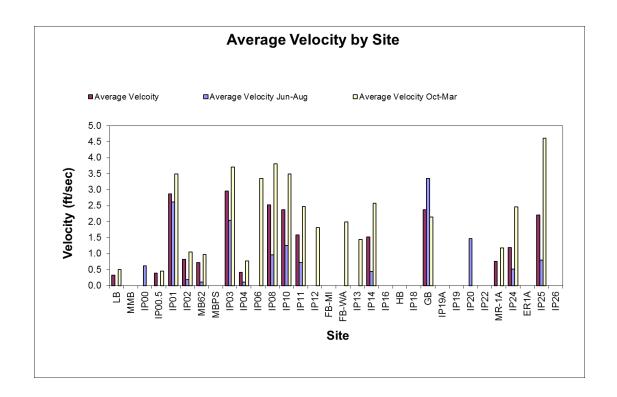


Figure 11: Comparison of average annual, spring and summer water depths by site.

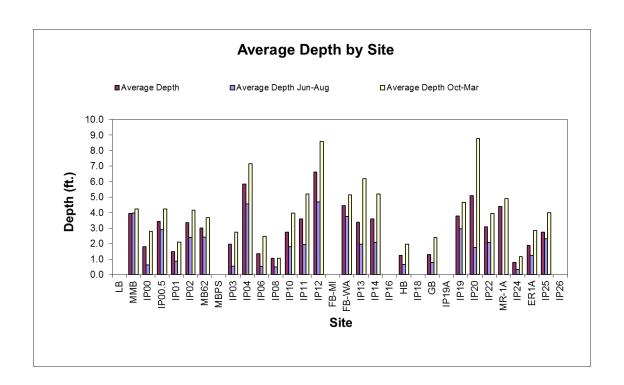
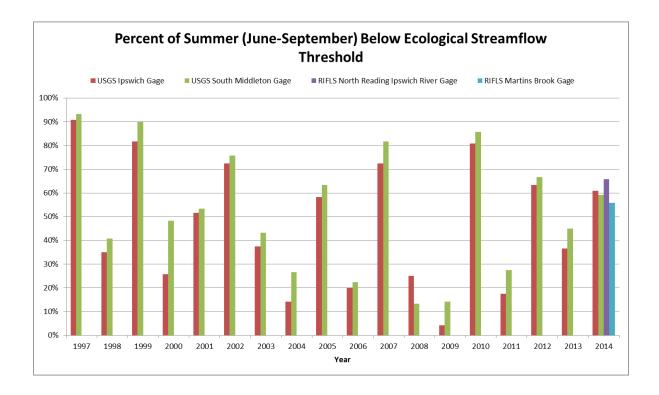


Figure 12. Percent of days during summer months (June-September) when flows fall below ecological streamflow threshold of 0.42 cfsm at flow monitoring sites.



General Findings

Withdrawals for drinking water are the primary cause of unnaturally low flows in the Ipswich River (Armstrong 2001, Zarrielo and Ries 2000). While it might be expected that low flows occur seasonally, the low flows observed in the Ipswich River are about 10% of what might be considered "natural." Due to low flows, the Ipswich River is classified as highly stressed by the MA Water Resources Commission (2001) and impaired under section 303(d) of the Clean Water Act.

Flow monitoring data indicate that fluctuations and differences in flows are more pronounced below the established threshold of 0.42 cfsm. Further analysis is needed to determine the exact role that groundwater withdrawals and land cover may have in causing the observed changes. Having access to continuous data at the RIFLS gages will also be important to determine the statistical significance of the observed trends. Water level loggers will continue to be used in 2015 at the Martins Brook and Haverhill St. locations.

Conductivity

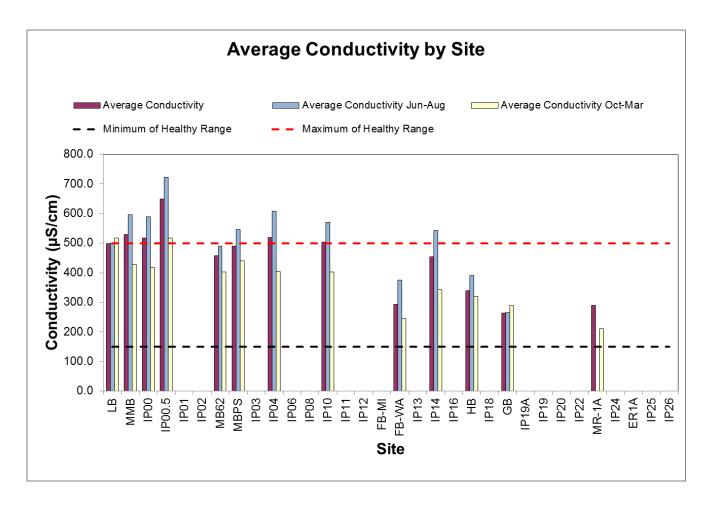
Conductivity measures the ability of water to pass an electrical current resulting from the presence of dissolved solids (or salts) such as chloride, sulfate, sodium and calcium, among others. Many factors can affect conductivity including local geology, rainfall, low flows and salt water concentrations in tidal areas. Most streams have a fairly constant range of conductivity under normal circumstances. Therefore, significant changes in conductivity can be an indicator that a discharge or some other source of pollution has entered the water. According to the EPA, the conductivity of rivers in the United States generally ranges from 50 to 1500 μ S/cm (micro Siemens per centimeter). Rivers that can support healthy fisheries should be in the range of 150 to 500 μ S/cm.

Conductivity was measured at 14 sites in 2014. Table 6 shows statistics of conductivity collected from 2007, when measurements began, through 2014. Figure 13 shows a comparison of average annual and summer conductivity for the sites monitored. Conductivity monitoring was expanded between 2013 and 2014. The increase in the types of sites monitored and the greater amount of data shows an increase in the percent of samples exceeding water quality guidelines from previous years. Many sites show higher readings in the winter months, particularly in the upper watershed. Roadside salt applications are known to increase chloride concentrations at wellfields located in the Ipswich River Watershed in Wilmington, MA (Heath, et al., 2012). Elevated conductivity values are often seen in the spring, especially at sites in the upper watershed, possibly due to the influx of road salts from stormwater runoff. These values are at or exceed those that are known to support healthy fisheries. Continuing to monitor conductivity will be important to establish baseline trends and observe regional differences.

Table 6: Annual statistics for conductivity.

| | | | | | Annual | Summer | | % Samples |
|--------------|------|-----------|---------|---------|-----------|---------|-------------|-----------------|
| | | # Samples | | | Average | Average | | Exceeding Water |
| | | (March- | | | (March- | (June- | # Samples > | Quality |
| Parameter | Year | December) | Minimum | Maximum | December) | August) | 500μS/cm | Recommendations |
| Conductivity | | | | | | | | |
| (μS/cm) | 1997 | | | | | | | |
| | 1998 | | | | | | | |
| | 1999 | | | | | | | |
| | 2000 | | | | | | | |
| | 2001 | | | | | | | |
| | 2002 | | | | | | | |
| | 2003 | | | | | | | |
| | 2004 | | | | | | | |
| | 2005 | | | | | | | |
| | 2006 | | | | | | | |
| | 2007 | | | | | | | |
| | 2008 | 27 | 150 | 517 | 352 | 319 | 0 | 0% |
| | 2009 | | | | | | | |
| | 2010 | | | | | | | |
| | 2011 | 37 | 180 | 620 | 395 | 414 | 3 | 8% |
| | 2012 | 79 | 170 | 610 | 424 | 454 | 6 | 8% |
| | 2013 | 79 | 200 | 840 | 469 | 425 | 8 | 10% |
| | 2014 | 101 | 200 | 770 | 472 | 538 | 22 | 22% |

Figure 13. Annual and summer average conductivity by site. The conductivity range considered suitable for healthy fisheries is 150-500 μ S/cm (micro Siemens per centimeter).



Color and Odor

The Ipswich River is a tea-like color naturally. This color is due primarily to dissolved organic carbon (e.g., tannins from leaves and plants). There is a lot of dissolved organic carbon in the Ipswich River due to the wetlands that drain into the river throughout the watershed.

Each month monitors noted the color and odor of the river on their data sheets in order to track changes or events where color changed significantly. Color was measured on a scale of 1 through 5: 1 (Clear), 2 (Very Light Tea), 3 (Light Tea), 4 (Tea), and 5 (Dark Tea). If a particular odor was noticed, this was noted on the data sheet. Most colors noted were in the Very Light Tea to Light Tea range. The river tended to be a light tea throughout the year.

Darker colors (tea to dark tea) were typically recorded in the summer months (July – August) and so may be associated with lower flow periods. However, in general it seems that there is no clear relationship between darker color and higher flow periods. Some sites were darker when it rained, some sites were variable, and some were lighter. It does seem, however, that darker colors were prevalent during summer months, and particularly associated with lower flows.

Habitat Observations

Each month monitors recorded wildlife and habitat observations. Often, the level of observation depended on monitor knowledge of birds, macroinvertebrates, fish, and other wildlife. Lists of birds and other wildlife seen are below.

Beaver activity was noted at sites IP01 and IP13. Fish activity was noted at IP10 and FB-MI.

Birds

Baltimore Oriole Downy Woodpecker Pine Siskins

Barn Swallows Eastern Wood Peewee Red-Bellied Woodpecker

Belted Kingfisher Goldfinch Red-Tailed Hawk

Blue Jay Grackles Robins
Broad Winged Hawk Great Blue Herron Rusty Crows
Canada Geese Great Crested Flycatcher Song Sparrow
Cardinals Greater Yellowlegs Starling

Cedar WaxwingHairy WoodpeckerSwamp SparrowChickadeesHerring GullTufted TitmouseChimney SwiftMallardsTurkey Vulture

Common Yellowthroat Mourning Dove White Breasted Nuthatch

Crows Mute Swan Wood Duck
Dark Eyed Juncos Baltimore Orioles Yellow Warbler

Double Crested Cormorant Phoebe

Other Wildlife

Mammals: Beavers, Muskrat, Mink

Reptiles and Amphibians: Frogs, Toads, Snakes, Painted Turtle, Snapping Turtles

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Insects: Dragonflies, crickets

<u>Plants</u> Loosestrife, duckweed, cattails, pickerel weed

Other Observations

| Site ID | Date(s) | Observation |
|---------|---------|-------------------------------|
| MMB | 7/24 | Odor gross! |
| | | Ammonia/metallic? Pickerel |
| | | weed is blooming. |
| MMB | 8/31/14 | River is gross! Brown/greasy |
| | | to the touch. Odor of "wet |
| | | feathers". |
| MMB | 10/24 | Water looks very dark, but |
| | | weirdly greenish, brownish in |
| | | bucket. |
| IP00 | 4/27 | Metallic odor |
| IP00 | 9/28 | River looks awful! NO FLOW |
| IP00.5 | 9/28 | River is lowest of the year |
| IP11 | 4/27 | Two fly fishermen caught 3 |
| | | rainbow trout. |

3.2 Quality Assurance/Quality Control

Quality Assurance Project Plan (QAPP)

A formal Quality Assurance Project Plan (QAPP) was updated and approved in April of 2013 for the RiverWatch Program by the Department of Environmental Protection (DEP) and the Office of Coastal Zone Management (CZM). No changes were made to the QUAPP for the 2014 monitoring year.

As part of the Quality Assurance Project Plan (QAPP), both temperature and dissolved oxygen are evaluated for quality control purposes. Volunteers must attend an annual training and calibrate new chemicals for testing dissolved oxygen. Also, monitors undergo an annual site audit by the Program Coordinator from IRWA where values for dissolved oxygen and temperature obtained by the program manager are compared to the values obtained by the monitor. Monitors also perform a duplicate test for dissolved oxygen once each year.

Three training events were held in 2014 to increase the likelihood of full attendance. All monitors attended except for one site (IP22). Site MMB is covered by the same volunteers who also test IP00.

Table 7 shows results for dissolved oxygen and temperature calibration values at the annual training. An optical dissolved oxygen probe and digital meter from Thermo Scientific was calibrated at air saturation and used to generate the calibration standard values for dissolved oxygen and temperature whereas the volunteers use Winkler Titration kits with new chemicals and standard thermometers. Thermometers are replaced on an as-needed basis. Water temperature readings would exceed the acceptable calibration range of 1.0 °C when the volunteer recorded their reading much later than others so that the water was warmer as confirmed by a second check with the meter.

Comparison of program manager site audit DO and temperature readings are presented in table 8. This type of field duplicate is performed by first calibrating the dissolved oxygen meter at air saturation and taking a reading from either the bucket grab sample or stream depending on where the volunteer fills the sample bottle to begin the Winkler Titration procedure. For 2014 site audit comparisons, 8 sites exceeded the 1 mg/L DO concentration and all temperature readings fell within the 1.0 °C difference QA/QC standard. Volunteers perform one field duplicate per year on their own in July and these results are presented in table 9. All but one of the field duplicates met quality standards, indicating that volunteer data are within quality assurance limits.

Table 7. Volunteer training dissolved oxygen and temperature calibration comparisons.

| Site | Date | Attended training | DO monitor | DO Trainer | Difference | Acceptable | Action Taken | Temp Monitor | Temp Trainer | Difference | Acceptable | Action |
|----------|-----------|-------------------|---------------|---------------|------------|------------|---|-----------------|-----------------|------------|------------|---|
| LB | 3/22/2014 | yes | 9.2 | 9.8 | 0.6 | yes | | | | 0 | yes | |
| MMB/IP00 | 3/15/2014 | Yes | 8.2 | 9.4 | | | | 14 | 15 | | | |
| | | | | | | | | | | | | Temperature measured later than |
| IP00.5 | 3/22/2014 | Yes | 10 | 9.8 | | yes | | 7.0 | 4 | | no | others |
| IP01 | 3/22/2014 | Yes | 10 | 9.8 | | yes | | 3.0 | 4 | | yes | |
| IP02 | 3/22/2014 | yes | 10 | 9.8 | 0.2 | yes | performed second | 3.0 | 4 | 1 | yes | |
| MB-PS | 3/22/2014 | Yes | 8.6 | 9.8 | 1.2 | no | titration | | | 0 | yes | |
| MB-62 | 3/13/2015 | Yes | 8.2 | 8.2 | 0 | yes | | 13.0 | 13 | | | |
| IP03 | 3/22/2014 | Yes | 9.8 | 9.8 | 0 | yes | | 3.0 | 4 | 1 | yes | |
| IP04 | | | | | | | | | | | | |
| IP06 | 3/15/2014 | Yes | 8.8 | 9.4 | 0.6 | yes | | | | 0 | yes | |
| IP08 | 3/15/2014 | Yes | 8 | 9.4 | 1.4 | no | performed second titration | 14.0 | 15 | 1 | yes | |
| IP10 | 15-Mar | Yes | 7.4 | 8 | | ves | | 15.0 | 15 | | yes | |
| IP11 | 3/22/2015 | Yes | 10 | 9.8 | | yes | | 5.0 | 4 | | yes | |
| IP12 | 3/15/2014 | Yes | 8.2 | 9.4 | 1.2 | | | 15.0 | 15 | | yes | |
| FB-WA | 3/15/2014 | Yes | 8.6 | 9.4 | | yes | | 15.0 | 15 | | ves | |
| FB-MI | 3/22/2015 | Yes | 9.2 | 9.8 | | yes | | 3.9 | 4 | | yes | |
| IP13 | 3/8/2015 | Yes | 8.3 | 8.3 | | ves | | 8.0 | 8 | | ves | |
| IP14 | 3/15/2014 | Yes | 8.8 | 9.4 | 0.6 | yes | performed | 13.0 | 15 | 2 | no | Temperature measured later than others |
| IP16 | 3/15/2014 | Yes | 7.8 | 9.4 | 1.6 | no | second titration | 15.0 | 15 | 0 | yes | |
| HB | 3/8/2014 | Yes | 7.8 | 8.3 | | yes | uuduon | 8.0 | 8 | | yes | |
| IP18 | 9,0,=011 | | | | | ,,,, | | | | | ,,,, | |
| GB | 3/8/2014 | yes | 7.2 | 8.3 | 1.1 | no | performed second titration performed | 8.0 | 8 | 0 | yes | |
| IP19/19A | 3/15/2014 | Yes | 8 | 9.4 | 1.4 | no | second titration | 14.5 | 15 | 0.5 | yes | |
| IP20 | 3/8/2014 | Yes | 7.9 | 8.3 | 0.4 | yes | | 8.0 | 8 | | yes | |
| IP22 | | | | | | | | | | | | |
| MR-1A | 3/8/2014 | Yes | 8.4 | 8 | 0.4 | yes | | 8.0 | 8 | 0 | yes | |
| IP24 | 3/8/2014 | Yes | 8.1 | 8.3 | 0.2 | yes | performed second | 8.0 | 8 | 0 | yes | |
| ER-1A | 3/8/2014 | Yes | 6.5 | 8.3 | 1.8 | no | titration | 9.0 | 8 | 1 | yes | |
| IP25 | 3/8/2014 | Yes | 7.4 | 8.3 | | yes | | 7.5 | 8 | | ves | |
| IP26 | 3/8/2014 | Yes | 8.2 | 8.3 | | ves | | 9.0 | 8 | | ves | |

Table 8. Program manager site audit comparisons for dissolved oxygen and temperature readings in 2014.

| Site | Date | Auditor DO | Monitor DO | Difference | Acceptable | Action Taken | Auditor Temp | Monitor Temp | Difference | Acceptible | Action Taken |
|----------|------------|------------|------------|------------|------------|--------------|--------------|--------------|------------|------------|--------------|
| LB | 5/18/2014 | 4.6 | 4.0 | 0.6 | yes | | 19 | 19 | 0.0 | yes | |
| MMB | | | | | | | | | 0.0 | yes | |
| IP00 | 5/18/2014 | 4.6 | 4.0 | 0.6 | yes | | 16 | 17 | 1.0 | yes | |
| IP00.5 | 6/29/2014 | 7.3 | 5.3 | 2.0 | no | | 22 | 23 | 1.0 | yes | |
| IP01 | 6/29/2014 | 3.1 | 2.7 | 0.4 | yes | | 21.5 | 21.9 | 0.4 | yes | |
| IP02 | | | | | | | | | 0.0 | yes | |
| MB-PS | 11/16/2014 | 10.6 | 9.2 | 1.4 | no | | 2 | 2.4 | 0.4 | yes | |
| MB-62 | 12/14/2014 | | | | | | 3 | 3.4 | 0.4 | yes | |
| IP03 | 12/14/2014 | 11.0 | 10.0 | 1.0 | yes | | 2.5 | 2.64 | 0.1 | yes | |
| IP04 | 11/16/2014 | 11.2 | 9.4 | 1.8 | no | | 2.7 | 2.9 | | yes | |
| IP06 | 11/16/2014 | 12.7 | 10.2 | 2.5 | no | | 3.5 | 3.3 | 0.2 | yes | |
| IP08 | 11/16/2014 | 10.3 | 9.3 | 1.0 | yes | | 2 | 2.5 | 0.5 | yes | |
| IP10 | 10/16/2014 | 5.5 | 4.0 | 1.5 | no | | 10.5 | 11.4 | | yes | |
| IP11 | 8/24/2014 | 6.7 | 6.3 | 0.4 | yes | | 20 | 20.7 | 0.7 | yes | |
| IP12 | 10/26/2014 | 6.5 | 6.6 | 0.1 | yes | | 11.1 | 11.8 | 0.7 | yes | |
| FB-WA | 3/30/2014 | 11.2 | 10.1 | 1.1 | no | | 2 | 3 | 1.0 | yes | |
| FB-MI | | | | | | | | | 0.0 | yes | |
| IP13 | 8/1/2414 | 6.7 | 6.8 | 0.1 | yes | | 19 | 19.5 | 0.5 | yes | |
| IP14 | 10/26/2014 | 6.8 | 6.2 | 0.6 | yes | | 11 | 11.6 | 0.6 | yes | |
| IP16 | 8/24/2014 | 1.8 | 1.5 | 0.3 | yes | | 20 | 20.5 | 0.5 | yes | |
| НВ | 10/26/2014 | 9.2 | 8.3 | 0.9 | yes | | 11 | 11.4 | 0.4 | yes | |
| IP18 | 7/27/2014 | 2.1 | 2.3 | 0.2 | yes | | 24 | 24 | 0.0 | yes | |
| GB | 7/27/2014 | 5.4 | 5.0 | 0.4 | yes | | 22 | 21.6 | 0.4 | yes | |
| IP19/19A | 7/27/2015 | 5.1 | 4.5 | 0.6 | yes | | 24 | 25 | 1.0 | yes | |
| IP20 | 4/27/2014 | 8.8 | 7.7 | | | | 10 | 10.2 | 0.2 | yes | |
| IP22 | 4/27/2014 | 9.4 | 8.9 | 0.5 | yes | | 10 | 10.4 | 0.4 | yes | |
| MR-1A | 3/30/2014 | 11.0 | 10.8 | 0.2 | yes | | 3.5 | 4 | | yes | |
| IP24 | 3/30/2014 | 11.0 | 10.0 | 1.0 | yes | | 4 | 4 | 0.0 | yes | |
| ER-1A | 4/17/2014 | 10.6 | 9.0 | | | | 11 | 10.9 | 0.1 | yes | |
| IP25 | | | | | | | | | 0.0 | yes | |
| IP26 | 11/16/2014 | 12.5 | 11.6 | 0.9 | yes | | 4.5 | 5.2 | 0.7 | yes | |

Table 9. Monitor field duplicate dissolved oxygen measurements for 2014.

| Site | Date | DO 1 | DO 2 | Difference | Acceptible |
|----------|-----------|------|------|------------|------------|
| LB | 7/27/2014 | 1.4 | 1.4 | 0.0 | yes |
| MMB | 7/27/2014 | 1.2 | 1.4 | 0.2 | yes |
| IP00 | 7/27/2014 | 1.0 | 1.6 | 0.6 | yes |
| IP00.5 | 7/27/2014 | 4.5 | 3.7 | 0.8 | yes |
| IP01 | 7/27/2014 | | | - | |
| IP02 | 7/27/2014 | | | - | |
| MB-PS | 7/27/2014 | 4.3 | 4.2 | 0.1 | yes |
| MB-62 | 7/27/2014 | | | - | |
| IP03 | 7/27/2014 | 3.6 | 3.8 | 0.2 | yes |
| IP04 | 7/27/2014 | 4.4 | 4.6 | 0.2 | yes |
| IP06 | 7/27/2014 | 6.2 | 6.2 | 0.0 | yes |
| IP08 | 7/27/2014 | 6.2 | 6.2 | 0.0 | yes |
| IP10 | 7/27/2014 | 2.4 | 3.0 | 0.6 | yes |
| IP11 | 7/27/2014 | 4.8 | 4.4 | 0.4 | yes |
| IP12 | 7/27/2014 | 5.2 | 5.5 | 0.3 | yes |
| FB-WA | 7/27/2014 | 5.0 | 5.2 | 0.2 | yes |
| FB-MI | 7/27/2014 | | | - | |
| IP13 | 7/27/2014 | 6.0 | 6.0 | 0.0 | yes |
| IP14 | 7/27/2014 | 4.6 | 4.8 | 0.2 | yes |
| IP16 | 7/27/2014 | | | - | |
| HB | 7/27/2014 | 4.1 | 4.3 | 0.2 | yes |
| IP18 | 7/27/2014 | | | - | |
| GB | 7/27/2014 | 5.0 | 4.1 | 0.9 | yes |
| IP19/19A | 7/27/2014 | 4.6 | 4.4 | 0.2 | yes |
| IP20 | 7/27/2014 | | | - | |
| IP22 | 7/27/2014 | 1.4 | 3.0 | 1.6 | no |
| MR-1A | 7/27/2014 | | | - | |
| IP24 | 7/27/2014 | 4.4 | 4.3 | 0.1 | yes |
| ER-1A | 7/27/2014 | 5.3 | 5.0 | 0.3 | yes |
| IP25 | 7/27/2014 | | | - | |
| IP26 | 7/27/2014 | 5.7 | 6.1 | 0.4 | yes |

Volunteer Qualifications

Volunteer quality assurance is maintained in the following ways:

Volunteers attend one training annually, led by the Monitoring Coordinator. The training includes a review of all procedures in the RiverWatch Monitoring Manual and a discussion of any changes. In addition, the previous year's data are presented, calibrations conducted, and QA/QC standards discussed.

Monitors are audited at their sampling site once per year.

Volunteers take duplicate samples at their site once per year, and equipment, data analysis and data control are held to QA/QC standards to the maximum extent possible.

Completeness

Table 10, below, summarizes the completeness of data collection for the 17-year period. Completeness is calculated as the number of samples taken in a year divided by the maximum number of samples it was possible to collect during that year. Our goal is to collect at least 80% of the total number of samples possible, and that goal was met for every year except 2003 and 2010. However, there is excellent completeness for all other years of monitoring, indicating the strength of volunteer commitment. In 2009, the bridge at site IP18 was out for construction, so monitoring was not possible for six months.

Table 10: Percent of Samples Collected per year, 1997 - 2014.

| Year | Completeness | Year | Completeness |
|------|--------------|------|--------------|
| 1997 | 86% | 2006 | 91% |
| 1998 | 90% | 2007 | 82% |
| 1999 | 92% | 2008 | 83% |
| 2000 | 89% | 2009 | 78% |
| 2001 | 83% | 2010 | 73% |
| 2002 | 89% | 2011 | 85% |
| 2003 | 76% | 2012 | 87% |
| 2004 | 81% | 2013 | 82% |
| 2005 | 88% | 2014 | 87% |

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RiverWatch Data Appendices and Quality Assurance Project Plan (QAPP) available at: http://www.ipswichriver.org/our-work/library/research/